#### 1 Discrete or indiscrete? Redefining the colour polymorphism of the land snail

#### 2 Cepaea nemoralis

- 3 Angus Davison\*, Hannah J. Jackson, Ellis W. Murphy, Tom Reader
- 4 School of Life Sciences, University of Nottingham, Nottingham NG7 2RD, United
- 5 Kingdom
- 6 \*E-mail: angus.davison@nottingham.ac.uk
- 7 Running head: Redefining a snail colour polymorphism

Biologists have long tried to describe and name the different phenotypes that make
up the exuberant colour polymorphism of the land snail *Cepaea nemoralis*.

Traditionally, the view is that the ground colour is one of a few major colour classes, 10 either yellow, pink or brown, but in practise it is frequently difficult to distinguish the 11 colours, and consistently define different shades of the same colour. To understand 12 whether colour variation is continuous, and to investigate how the variation may be 13 14 perceived by an avian predator, we applied psychophysical models of colour vision to shell reflectance measures. The main finding is that both achromatic and 15 16 chromatic variation are indiscrete, being continuously distributed over many perceptual units, with the major axis of chromatic variation representing differences 17 in saturation, or purity of colour. Nonetheless, clustering analysis based on the 18 density of the distribution revealed three groups, roughly corresponding to human-19 perceived yellow, pink and brown shells. There is also large-scale geographic 20 variation between these morphs across Europe, and some covariance between shell 21 colour and banding patterns. Although further studies are necessary to understand 22 the evolutionary origins and impact of natural selective upon this variation, the 23 observation of continuous variation in colour is intriguing, given that the underlying 24 supergene that determines colour should prevent phenotypes from "dissolving" into 25 continuous trait distributions. 26

27 Keywords: Cepaea, colour, polymorphism, snail, supergene

Throughout the past century, the study of animal colour has been critical in making 28 progress in understanding the principles of biology, especially with respect to 29 genetics and evolution (McKinnon and Pierotti 2010; McLean and Stuart-Fox 2014; 30 Cuthill et al. 2017; San-Jose and Roulin 2017). For instance, early studies on the 31 inheritance of colour traits were important in establishing an understanding of basic 32 Mendelian genetics (Wheldale 1907; Staples-Browne 1908). Subsequently, studies 33 34 of the distribution and predation of colour morphs have and continue to shape our understanding of how natural and sexual selection operate in wild populations 35 36 (Hugall and Stuart-Fox 2012; Dale et al. 2015; Delhey et al. 2017). Most recently, candidate gene and latterly genomic approaches have been used to identify the 37 underlying genes that determine the colour differences (references in Hoekstra 2006; 38 McLean and Stuart-Fox 2014; San-Jose and Roulin 2017). 39

For practical reasons, many of these prior studies have taken advantage of traits that exhibit relatively simple, discrete variation and straightforward inheritance patterns, but this risks missing the extraordinary variation of life forms and colour traits. It is also likely that in nature discrete variation is the exception rather than the rule – and this is becoming more evident as researchers increasingly use instrumentation to measure colour (Montgomerie 2006), rather than being obliged to bin types into human-defined categories.

Historically, two of the most important animals in studying colour
polymorphism have been the peppered moth *Biston betularia* and the grove snail *Cepaea nemoralis* and its sister taxon, *C. hortensis* (collectively "*Cepaea*", the
preferred common name), because individuals are relatively easy to collect and
study, and the colour morphs show straightforward inheritance. However, while
ongoing and long-term studies on these animals continue to provide compelling

evidence for the fundamental role of natural selection in promoting and maintaining
variation in natural populations, as well as the impact of modern-day habitat change
(Silvertown et al. 2011; Cook et al. 2012), progress in understanding the precise
mechanism of the polymorphisms has diverged in the two systems.

57 In the peppered moth, the precise mutation that determines the colour 58 differences was reported to be in a known patterning gene (van't Hof et al. 2016). In contrast, an understanding of the 'exuberant' (Franks and Oxford 2009) 59 polymorphism of Cepaea – whether yellow, pink or brown shells and zero to five 60 bands – has stalled since the 1970s. In part, this may be a reaction to the question of 61 Jones et al. (1977) on whether the Cepaea polymorphism is "a problem with too 62 many solutions?" Actually, the intention of that work was to emphasise the perfect 63 case study provided by Cepaea; as simple explanations for phenotypic variation are 64 65 the exception, Jones et al. were making the point that it is important to study organisms for which polymorphism may be explained by a variety of processes. 66 precisely because they are more realistic. Some 40 years later, this comment is still 67 relevant, given that genomic technologies and DNA sequence analyses should allow 68 us to uncover the relative contributions of each of these processes to contemporary 69 diversity - but it is nonetheless understandable that most study systems are still 70 selected because of their relative simplicity. 71

Despite a general lack of progress, the *Cepaea* polymorphism retains excellent potential as a model system in evolutionary biology. Previous studies have laid the foundations for future progress, including exploiting many long term studies (Cook et al. 1999; Davison and Clarke 2000; Silvertown et al. 2011; Cameron and Cook 2012; Ożgo and Schilthuizen 2012; Cameron et al. 2013; Schilthuizen 2013; Cook 2014; Ozgo et al. 2017), increasing understanding of the pigments and shell

proteome (Mann and Jackson 2014; Williams 2017) and especially, using new 78 genomic methods to identify the genes involved (Richards et al. 2013; Kerkvliet et al. 79 2017). Of particular interest here, we note (as do others, Surmacki et al. 2013) that 80 there is now a pressing need to quantify objectively the polymorphism of Cepaea 81 shells, and to understand how this is perceived by an avian predator, because only 82 then can we properly understand how the polymorphism is maintained. The specific 83 84 problem is that in the past Cepaea shell colours have usually been treated as one of three or more distinct classes (e.g. Cain and Sheppard 1954) – vellow, pink or brown 85 86 - partly due to a lack of objective measures of colour, especially those that can be used in the field and between different observers and contexts (Cain et al. 1960; 87 Cain et al. 1968; Jones et al. 1977). There has been also the significant issue that 88 human perception of colour is not necessarily objective or the same as that of an 89 avian predator (Surmacki et al. 2013). 90

91 Now that objective methods of measuring and analysing colour are widely available (Endler 1990; Montgomerie 2006; Maia et al. 2013; Delhey et al. 2015) and 92 93 easy to use, at least in a controlled laboratory setting, we set out to measure guantitatively the ground colour of *Cepaea* shells, and so to define the nature of the 94 polymorphism. Specifically, by measuring the shell colour of snails collected across 95 the breadth of the European distribution, we used psychophysical models of colour 96 vision to assess how chromatic variation is perceived by birds (but not categorised: 97 Caves et al. 2018). 98

99 Previously, Surmacki *et al.* (2013) used quantitative measures of colour on 100 relatively few individuals to assess how shells match to various backgrounds. Here, 101 we investigate the extent to which the distribution of snail shell colour is continuous 102 along the main axes of chromatic variation, using more than a 1000 individuals and

Gaussian finite mixture modelling (Scrucca et al. 2016) to test whether colours fall
into clusters in multivariate space. We also aimed to understand if quantitative
measures on a relatively small sample of shells can describe – rather than explain –
geographic patterns in colour morph frequency across Europe, as others have done
in much larger qualitative surveys.

The findings have significance for understanding the *Cepaea* polymorphism, and the nature of the selection that acts upon it, as well as more generally highlighting the need to measure colour objectively in other systems, before being able to test for possible explanations.

# 112 Methods

#### 113 Data collection

Individual C. nemoralis snails were mainly gathered opportunistically by volunteer-114 led collection and field trips across Europe (Grindon and Davison 2013). Snails were 115 frozen upon arrival at the University of Nottingham, subsequently thawed and the 116 117 body extracted from the shell. The ground colour and banding of the shell was then scored qualitatively by an experienced person (A.D.) and a student, as either yellow 118 (Y), pink (P) or brown (B), and unbanded (O), mid-banded (M), or all other banding 119 patterns (B, usually five-banded). Subsequent statistical analyses were carried out at 120 the level of the individual and the level of the population (sample site). So that we 121 122 could compare broad-scale patterns across Europe, larger groups were also used – individual sample sites were therefore grouped into one of six larger groups (Table 123 S1; Figure 1). 124

125 An Ocean Optics spectrometer (model USB2000+UV-VIS-ES) and a light 126 source (DT-MINI-2-GS UV-VIS-NIR) were used to measure individual reflectance

spectra of shells, using a WS-1 diffuse white reflectance standard to set the baseline 127 light spectrum (Teasdale et al. 2013; Taylor et al. 2016), and complete darkness to 128 set the dark spectrum standard. Reflectance measurements were taken on the 129 underside of the dried shell, because it was usually the least damaged region, least 130 exposed to sunlight, and well away from any bands. Point samples were taken for 131 each shell at a 45° incident angle, 2 mm from the shell. Individual shells were 132 133 measured three times, non-consecutively, with the software recalibrated against light standards every 2-5 measurements. Readings were collected using Ocean Optics 134 135 SpectraSuite v. 2.0.162 (software settings: integration time 750msec, boscar width 5, scans to average 10); then the raw data smoothed and binned into 5 nm categories 136 using Pavo version 0.5-6 (Maia et al. 2013). 137

138 Analysing chromatic and achromatic variation

We used the framework provided by Delhey et al. (2015) to analyse the reflectance 139 140 spectra. In this framework, a psychophysical model of colour vision (Vorobyev and Osorio 1998; Vorobyev et al. 1998) is used to assess whether chromatic differences 141 between reflectance spectra exceed a discrimination threshold, or 'just noticeable 142 difference' (JND), which can be perceived by a receiver, such as an avian predator. 143 The key to these models lies with the degree to which a particular combination of 144 reflectance and illuminant spectra stimulate each of the different photoreceptors in 145 the retina. In birds, these photoreceptors are the four single cones used for colour 146 vision, which are sensitive to long (L), medium (M), short (S), and very short (VS) 147 148 wavelengths of light (Cuthill 2006).

To analyse chromatic variation, the quantum catches for each cone type were converted into three chromatic coordinates (x, y and z), where Euclidean distances between points reflect perceptual differences, using the formulas of Cassey *et al.* 

(2008). As there are no data for the song thrush, *Turdus philomelos*, which is the 152 main predator of *Cepaea*, we used sensitivity functions for the closest available 153 relative, the blackbird Turdus merula (Hart et al. 2000; Hart 2001), namely cone 154 proportions of VS: 0.528, S: 0.904, M: 1.128, and L: 1, sensitivity functions of 373, 155 461, 543 and 603, respectively. The analysis assumed that the L cone has a noise-156 to-signal ratio of 0.05, so that the ratios for the other cones were VS: 0.0688, S: 157 158 0.0526 and M: 0.0471. The irradiance spectrum of 'standard daylight' (d65) was used for the main analyses. However, analyses were also run for 'woodland shade' 159 160 to understand the influence of illuminant on avian perception of colour (Vorobyev et al. 1998). 161

To identify the main axes of chromatic variation, we carried out a Principal 162 Components Analysis (PCA) on the chromatic coordinates (x, y and z), preserving 163 the perceptual distances (JNDs) by using a covariance matrix rather than a 164 correlation matrix (Delhey et al. 2015). To understand whether there are potential 165 clusters within the chromatic coordinate data, Gaussian mixture modelling was 166 carried out using Mclust 5.3 in R version 3.3.3 (Scrucca et al. 2016). A number of 167 models were compared, each of which assumed a different number of clusters (from 168 1 to 10), normally distributed in multivariate chromatic space. Several classes of 169 model were considered, each with a different assumption about the homogeneity of 170 variance and orientation among clusters. The best fitting model was then determined 171 as the one with highest Bayesian Information Criteria (BIC), with significant 172 differences determined using a bootstrap approach. 173

The methods of Delhey et al. (2015) were also used to assess achromatic variation. In birds, sensitivity to achromatic cues is supposed to be mediated by double cones which have the same pigment as L cones in birds but different oil

droplets, so have a wider sensitivity range. Values of achromatic contrast were
therefore estimated, again in JNDs, by computing achromatic contrast between each
reflectance spectrum and a reference (a very low value of double cone quantum
catch, 0.001), corresponding to a dark spectrum, and using the same noise-to-signal
ratio.

182 Analysis of morph frequencies

We investigated evidence for effects of location and banding on the likelihood that a 183 snail belonged to a particular morph, using generalised linear mixed effects models 184 (GLMMs) with binomial errors. Each morph was considered separately, with each 185 snail to be scored as belonging to the focal morph (1) or not (0). The three analyses 186 are not independent, since each snail can only belong to one morph. Banding 187 pattern was fitted as a fixed factor, whilst the effect of geographic location was 188 examined at three spatial scales. Variation in morph frequency at a local level was 189 190 modelled with random effect for site. Variation at a regional level was considered by fitting a fixed effect of geographic region. Finally, continental scale variation was 191 modelled by looking for fixed linear and quadratic effects of latitude and longitude. 192 The fact that region and latitude/longitude are partially collinear was reflected in the 193 model-fitting procedure. We first fitted a saturated model with all main effects, except 194 for region, and their two-way interactions (excluding interactions involving quadratic 195 effects). Then, fixed terms were removed in a stepwise fashion, testing the effect of 196 deletion using likelihood ratio tests, until only significant terms remained. Effects of 197 198 latitude/longitude were then substituted with an effect of region and we compared the Akaike Information Criterion (AIC) of the resulting models, to test if region was better 199 200 at capturing any large-scale geographic variation. Testing random effects in 201 generalised linear mixed models is problematic, so we compared the AIC of the

saturated GLMM with that of a generalised linear model without the random term forsite to provide an approximate test of the importance of site.

## 204 **Results**

205 Variation in colour

We measured the individual reflectance spectra of 1172 shells, mainly collected from
across Europe (Table S1; Figure 1) and then transformed them into visual space
coordinates, xyz. To visualise this chromatic variation, and the relationship with
human-scored and Mclust-defined colour categories (below), the xyz coordinates
were plotted in visual colour space (Figure 2). There were no obviously discrete
groups.

A PCA on the xyz coordinates showed a first axis which explained 87% of 212 213 chromatic variation. PC1 had a moderate positive loading for x (0.61), and a moderate negative loading for v (-0.64) and z (-0.46). Two further axes explained 214 11% and 2% of the variation, the second having a positive loading on all axes (0.75, 215 216 0.28, 0.61, respectively), and the third a mixture (-0.26, -0.71, 0.65). The range of observed variation on each axis was considerable: 41, 22 and 8 JNDs for x, y and z 217 respectively (Figure 2). Plotting the average normalized reflectance spectra for each 218 guartile of each principal component showed how the three PC axes correspond to 219 chromatic variation (Figure 3). Variation along PC1 represents relatively high 220 221 stimulation of L cones and lesser stimulation of S cones, relative to M cones. Variation in PC2 showed relatively higher stimulation of L cones and lesser 222 stimulation of M cones. PC3 showed relatively high stimulation of the M cones 223 224 compared to lesser stimulation of the S and L cones. Only PC1 showed any differences in the VS region but the shells barely reflected in the UV. 225

To investigate whether snail shells cluster in chromatic space, and whether 226 observed clusters correspond to human-scored gualitative colour morphs, Gaussian 227 finite mixture modelling was applied to the xyz visual space coordinates. The best 228 229 model (VVV, ellipsoidal, varying volume, shape, and orientation; BIC -15727.3; P < 0.001 compared 2<sup>nd</sup> best model) recovered three clusters, roughly corresponding to 230 human-scored yellow (46%, n=539), pink (44%, n=511) and brown (10%, n=122) 231 (Table 1). The next best fitting model also recovered three clusters (VEV; BIC -232 15749.3; P < 0.001 compared with 3<sup>rd</sup> best model) and the third recovered four 233 clusters (EEV; BIC -15755.5; the 4<sup>th</sup> cluster contained only 16 individuals). 234 Comparing human-scored (A.D.) and Mclust-defined groups, the overall 235 concordance was good at 76% (Table 1), with a similar error rate (75%) for the 236 student group. The highest proportion of discordant scores were human-scored 237 yellow shells that Mclust classed as pink (10% for A.D.; 12% for student group), with 238 the other major discrepancies being human-pink classed as Mclust-brown (8%), and 239 human-yellow classed as Mclust-pink (4%). The main difference was that human-240 241 scoring reported relatively few brown shells (n=37), whereas the same group in Mclust was larger (n=122). With misclassifications adjusted relative to the total 242 number of each Mclust shell type, 81% of the brown group were in a different 243 human-scored group (74% pink, 7% yellow), compared to 26% of the pinks (3% 244 brown 23% vellow) and just 8% of vellows (8% pink, 0% brown). Thus, while the 245 overall correspondence between human and Mclust scoring of shell colour was 246 good, the yellows were scored accurately (92%), pinks less so (74%) and brown 247 poorly (19%). 248

Plots of human-scored colours along the three PCs (Supplementary Movie 1)
 and Mclust-categories were concordant with the above analyses (Figures 2, 4;

Supplementary Movie 2). Broadly, PC1 did not separate different human-perceived 251 colours or categories of shell, but instead mainly represents differences in saturation, 252 or purity of colour, between individuals (Figure 4). PC2 separated brown from yellow 253 and pink, and PC3 broadly separated all three colours, yellow, pink and brown. 254 255 The above analyses were repeated using woodland shade rather than 256 standard daylight. The main difference was that while Mclust again recovered three groups, brown shells were more common (14%, n=168), with fewer pinks (40%, 257 n=474) and approximately the same number of yellows (45%, n=530). 258 Finally, achromatic variation was also considerable, varying over more than 259 100 JNDs, and without any obvious differences between Mclust-defined colour 260 morphs (Supplementary Figure 1). 261 Geographic variation between morphs 262 Large-scale geographic variables (latitude, longitude and region) had significant 263 effects on the probability that a snail was pink or yellow, but not the probability that a 264 snail was brown (Table 2). Pink morphs were significantly less common at mid-265 266 latitudes (Figure 5). Snails with more than one band (B) and those which were mid-267 banded (M) were more likely to be pink in the west, while unbanded (O) snails were more likely to be pink in the east (Figure 5). In contrast, yellow snails were less 268 269 common at high latitudes, and were affected by an interaction between longitude and banding which was the reciprocal of that seen in pink snails (Figure 6). Morph 270 frequencies also varied at a local level: a saturated mixed model including the 271 272 random effect of site was much better (Brown AIC = 443.7; Pink AIC = 868.9; Yellow AIC = 852.8) than an equivalent model without the random effect (Brown AIC = 273 646.4; Pink AIC = 1407.9; Yellow AIC = 1416.2). Banding was associated with colour 274

morph in various ways. In addition to the interaction between banding and longitude
in pink and yellow snails mentioned above, unbanded snails (O) were generally more
likely to be brown (14% of all unbanded snails), than snails that were mid-banded
(M; 11.6%) or had several bands (B; 5.8%).

# 279 Discussion

By measuring the ground colour of Cepaea nemoralis shells collected across the 280 281 breadth of the European distribution, we used psychophysical models of avian vision to understand how the shell colour may be perceived by birds, and to describe how 282 this varies in geographic space, and with respect to other characters such as 283 284 banding. The findings have significance for understanding the Cepaea polymorphism, and the nature of the selection that acts upon it, as well as more 285 generally highlighting the need to objectively measure colour variation in other 286 systems before beginning to test for possible explanations. 287

Broadly, we found that both chromatic (Figure 2) and achromatic variation 288 (Supplementary Figure 1) is considerable, occurring over many perceptual units 289 (JNDs). If this variation, both within and among human-perceived colour morphs, 290 affects prey detection or identification by avian predators, then the presumption is 291 that the polymorphism must be impacted by natural selection. The current available 292 293 evidence suggests that animals in general use chromatic and achromatic signals for separate tasks, for example, using achromatic signals to identify the location, shape 294 and motion of objects, while chromatic signals identify surface quality (Osorio and 295 Vorobyev 2005). However, while this is also likely the case for avian predators, 296 specific experimental evidence from birds is sparse (Osorio et al. 1999; Kang et al. 297 2015; White and Kemp 2016). 298

In our analysis, we found that chromatic variation in shells is continuously
distributed in visual space, meaning that there are no wholly discrete colours (Figure
2). Perhaps surprisingly, we found that the most variable chromatic axis (PC1; 87%)
that would be visible to a bird reflects the degree of saturation, or purity of colour.
Axes separating human-perceived colours showed less variation, PC2 (11%)
separating brown from yellow/pink, and PC3 (2%) broadly separating yellow, pink
and brown.

306 Despite the lack of discrete colours, density-based clustering recovered three main shell types, which roughly correspond to human-perceived yellow, pink and 307 brown (Table 1; Figure 3). Brown shells were more common according to the 308 objective analysis than perceived by humans, with the frequency higher again when 309 using woodland shade as an illuminant. Therefore, prior studies that (necessarily) 310 used changes in frequencies of human-perceived colours to understand natural 311 selection on snail shells may have missed a significant part of the picture – not only 312 may birds use both achromatic and chromatic cues to differentiate morphs, but they 313 should also be able to perceive chromatic differences to a much finer precision than 314 a simple trivariate yellow, pink or brown categorisation that humans are obliged to 315 use in gualitative surveys. Of course, this does not mean that birds react to the many 316 morphs equally - it is possible that they categorically perceive a continuous variable 317 (Caves et al. 2018). Further investigations are needed, especially using a bird such 318 as the song thrush. 319

The effects of geographic location and banding pattern on variation in the reflectance spectrum of snails were also examined, the initial aim being to develop methods to *describe* variation, rather to *explain* it (e.g. by looking for correlations with environmental variables, putative selective agents, etc., as others have done;

Silvertown et al. 2011). Generally, we found that geographic variables (latitude, 324 longitude and region) and banding are generally associated with different 325 frequencies of the three traditional colour morphs, with the main directional trend 326 being that yellow snails are most common at mid-latitudes, as was found in much 327 larger studies (Jones et al. 1977; Silvertown et al. 2011). Similarly, as previously 328 reported (Cain et al. 1960), epistasis meant that unbanded snails (O) were generally 329 330 more likely to be brown, and banded snails (B) were less likely to be brown. Therefore, by establishing a method for quantitatively measuring colour, and 331 332 showing that a relatively small sample can be used to infer wide geographic patterns, this work provides a baseline for further studies on the polymorphism. 333 Discrete or indiscrete? 334 Laboratory crosses in the past have revealed that the variation in the Cepaea shell 335 phenotype is predominantly controlled by a 'supergene', which in a recent definition 336 337 is a genetic architecture involving multiple linked functional genetic elements that allows switching between discrete, complex phenotypes maintained in a stable local 338 *polymorphism* (Thompson and Jiggins 2014; Llaurens et al. 2017). This meaning fits 339

with the traditional view – and the classical 'Fordian' theory of polymorphism (Ford

341

342

1964) – that the ground colour of the shell is one of three more or less discrete

colour classes, either yellow, pink or brown, and indeed, is part of the reason that

343 *Cepaea* snails became a well-studied system. However, while scoring the shell 344 colour into different, discrete types is straightforward in offspring of individual crosses 345 in the lab, the acknowledged reality is that it is sometimes difficult to classify shells 346 consistently (e.g. see Table 1), especially when they are apparently intermediate in 347 form.

Now, in our study, we have shown definitively that the colour polymorphism of 348 Cepaea nemoralis is not discrete (Figures 2, 4). This finding emphasises the specific 349 practical problem for projects collecting and using shell polymorphism data, 350 especially those based entirely in the field and using citizen science (e.g. Evolution 351 Megalab; Silvertown et al. 2011). However, it also illustrates a more general 352 problem: if we do not have a precise definition of the Cepaea polymorphism and an 353 354 understanding of the underlying genetic control, then how can we claim to understand the evolutionary and ecological factors that maintain colour variation? 355 Mathematical modelling is one method that can be used to explore the 356 357 evolution of polymorphism, and of most relevance to this work, the circumstances that may or may not lead to discrete phenotypes. Historically, the argument of Ford 358 was that despite the fact that supergenes may appear as Mendelian loci, they were 359 actually rather complicated arrangements of several loci that are effectively 360 prevented from being broken up by recombination under most normal 361 circumstances. Thus, in both colour polymorphism in general (e.g. in side-blotched 362 lizards; Sinervo and Lively 1996) and specifically relating to supergenes (e.g. in 363 butterfly mimicry rings; Joron et al. 2011; Kunte et al. 2014), the distinctiveness of 364 the morphs is a central feature of the genetic control; the genetic architecture 365 specifically prevents phenotypes from "dissolving" into continuous trait distributions 366

367 (Ford 1964).

Much of the existing research has therefore begun from the premise of understanding how evidently discrete types come about, and thus give insight into the adaptive evolution of genome structure (Cuthill et al. 2017). In simulations it has been have shown that natural selection tends to favour lowered recombination when intermediate genotypes are at a disadvantage; unlinked loci modify the phenotype to

adapt to local conditions (e.g. to a local Batesian model butterfly; Charlesworth and
Charlesworth 1975b, a; Llaurens et al. 2017). More recently, and perhaps most
directly relevant to understanding the *Cepaea* polymorphism, Kopp and Hermisson
(2006) devised a model to investigate the evolution of a quantitative trait under
frequency-dependent disruptive selection. Their finding was that over generations
most of the genetic variation tends to concentrate on a small number of loci.

The historic background is perhaps part of the reason that most of the recent 379 progress in understanding supergenes has mainly come from species or systems 380 that show simple, wholly discrete phenotypes, for example in butterfly mimicry rings 381 (Joron et al. 2011), or heterostylous plants (Li et al. 2016). However, in Cepaea there 382 are many colour morphs, such that colour variation is quantitative and due to a 383 supergene; in other species such as the guppy *Poecilia* and the cichlid 384 385 Labeotropheus, the inheritance of often considerable colour variation is due to several loci, some sex-linked and others not (Tripathi et al. 2009; Thompson and 386 Jiggins 2014; Wellenreuther et al. 2014). Thus, developing theory on the impact of 387 negative frequency-dependent (apostatic) selection must be able to account for 388 these complexities, including those where supergenes are absent and variation is 389 quantitative, otherwise there is a risk that models will simply reaffirm what we already 390 know. 391

In one recent model, it was shown that crypsis and apostatic selection together may act to maintain a large number of morphs within a population, and in another apostatic selection was shown to maintain variation between similar species (Franks and Oxford 2011, 2017). In another more recent study, a simulation was used to explore the influence of predator perspective, selection, migration, and genetic linkage on colour allele frequencies. The relative sizes of predator and prey

home ranges can result in large differences in morph composition between 398 neighbouring populations (Holmes et al. 2017). Finally, in an empirical study blue 399 jays Cyanocitta cristata searched for digital moths on mixtures of dark and light 400 patches at different scales of heterogeneity. It was found that complex backgrounds 401 with many moth-like features elicited a slow, serial search that depended heavily on 402 403 selective attention. The result was increased apostatic selection, producing a broad 404 range of moth phenotypes (Bond and Kamil 2006). All of these circumstances may apply to the Cepaea colour polymorphism. 405

Overall, there is an open debate – but little empirical data – on how the 406 407 relative heterogeneity of the environment/substrate, density, distance or motion may influence the selection for crypsis or negative frequency dependence (Cuthill et al. 408 2017; Barnett et al. 2018). As Surmacki et al. (2013) summarised, if heterogeneous 409 areas consist of large patches of diverse habitats then this may promote the 410 evolution of specialist morphs through selection for crypsis, producing a few distinct 411 or specialist morphs, each more or less well matched to the coloration of the 412 preferred habitat type (Endler 1978; Bond 2007). If instead there are a mixture of 413 small microhabitats, apostatic selection is more likely to result in multiple morphs that 414 may be equally cryptic in all "grains" of the habitat. This is because in such 415 circumstances, predators use search images of the most common morph, and this 416 can lead to frequency-dependent selection. 417

418 Supergenes return

In contrast to a relative paucity of field data, and a relatively lack of progress in
establishing baseline theory, advances in DNA sequencing technology have meant
that knowledge on the genetics of colour polymorphism is advancing rapidly. As
hypothesised, in the still relatively few supergenes that have been fully

characterised, the discrete phenotypes are maintained due to close physical
proximity of the gene(s) and/or tight linkage (Joron et al. 2011; Kunte et al. 2014;
Gautier et al. 2018).

In comparison, a few more general studies on colour polymorphism, rather 426 than on supergenes specifically, have begun to reveal the extent of phenotypic 427 428 variation, and whether discrete or indiscrete. For example, reflectance spectra have been used to show that even though humans perceived the colour variation in the 429 eggs of African cuckoo finch Anomalospiza imberbis as falling into discrete 430 categories, the variation was actually continuous (Spottiswoode and Stevens 2010, 431 2011). Similarly, tawny dragon lizard Ctenophorus descresii does have discrete 432 colour morphs, but there is still considerable variation within each morph (Teasdale 433 et al. 2013). Further quantitative studies in other lizards in which colour 434 polymorphism has traditionally been treated as qualitative are also increasingly 435 showing that there are significant overlaps in colour (Cote et al. 2008; Vercken et al. 436 2008; Paterson and Blouin-Demers 2017). 437

In Cepaea nemoralis snails, the colour and banding elements of the 438 supergene have been mapped (Richards et al. 2013) but we remain ignorant of the 439 underlying genetics and the precise nature of the selection that acts upon the 440 polymorphism. For instance, models of supergene evolution require that intermediate 441 phenotypes are disadvantaged – this makes sense with respect to Batesian mimics 442 or distylous flowers – but in snails a rare intermediate might be at an advantage, due 443 444 to apostatic selection. At the molecular level, one scenario is that the extreme and effectively continuous colour variation of the shells is due to a corresponding high 445 number of colour alleles within the supergene. An alternative scenario is that there 446 447 actually relatively few colour alleles, with much of the chromatic variation due to

effects of other modifying loci (Charlesworth and Charlesworth 1975b). A final
consideration is that while colour variation might be continuous across a grand
geographic scale, if most local populations are founded by few individuals, then local
variation might be discrete, which is all that matters from a selective point of view.
This is more likely to be the case when both colour and banding are considered as
the visible phenotype, especially since they are frequently in linkage disequilibrium
(Cook 2017).

Overall, by establishing a method for quantitatively measuring colour, this 455 work provides a baseline for further studies on the polymorphism, both from the 456 perspective of understanding the nature of selection, and ultimately, also the genes 457 involved. To reconcile and test competing theories with the empirical observations, 458 the next steps must be to identify the component parts and evolutionary origins of 459 460 the supergene in *C. nemoralis*, develop a model of frequency-dependent selection, and further understand how birds react to specific elements of the chromatic 461 variation. When all of these findings are brought together, only then can we begin to 462 understand the evolutionary and ecological factors that maintain this "problem with 463 too many solutions." Whatever the final outcome, there is no risk that Cepaea snails 464 will be relegated to "other adaptive polymorphism" (Thompson and Jiggins 2014), 465 especially because, as Jones et al. (1977) suggested, it is important to study 466 organisms for which polymorphism may be explained by a variety of processes. 467 precisely because they are more realistic. 468

469 Supplementary material

470 Supplementary Movies 1, 2. Supplementary Figure 1. Supplementary Table 1.

471 Funding

- This work was mainly funded by the University of Nottingham with
- spectrophotometer purchased on the UoN equipment fund. Hannah Jackson is
- 474 funded by a BBSRC studentship.
- 475 Acknowledgements
- 476 Thanks to both Alan Bond and Laurence Cook for helpful discussions, and to and
- 477 Kaspar Delhey for discussion and assistance with the methods, to Alice Maiden and
- 478 Shagufta Hadife for collecting data, and to Adele Grindon and a network of helpers
- 479 collected the snails.
- 480 Data archiving
- 481 Raw reflectance data will be included with the manuscript or uploaded to Dryad upon
- 482 acceptance of the manuscript.

## 483 Literature cited

- Barnett, J. B., C. Michalis, N. E. Scott-Samuel, and I. C. Cuthill. 2018. Distance dependent defensive coloration in the poison frog *Dendrobates tinctorius*,
   Dendrobatidae. Proc Natl Acad Sci USA 115:6416-6421.
- Bond, A. B. 2007. The evolution of color polymorphism: crypticity searching images,
   and apostatic selection. Annu Rev Ecol Evol Syst 38:489-514.
- Bond, A. B. and A. C. Kamil. 1998. Apostatic selection by blue jays produces
   balanced polymorphism in virtual prey. Nature 395:594-596.
- Bond, A. B. and A. C. Kamil. 2002. Visual predators select for crypticity and
   polymorphism in virtual prey. Nature 415:609-613.
- Bond, A. B. and A. C. Kamil. 2006. Spatial heterogeneity, predator cognition, and the
  evolution of color polymorphism in virtual prey. Proc Natl Acad Sci USA
  103:3214-3219.
- Cain, A. J., J. M. B. King, and P. M. Sheppard. 1960. New data on the genetics of
   polymorphism in the snail *Cepaea nemoralis* L. Genetics 45:393-411.
- Cain, A. J. and P. M. Sheppard. 1954. Natural selection in *Cepaea*. Genetics 39:89 116.

- Cain, A. J., P. M. Sheppard, and J. M. B. King. 1968. Studies on *Cepaea*. I. Genetics
   of some morphs and varieties of *Cepaea nemoralis* (L). Philosophical
   Transactions of the Royal Society of London Series B-Biological Sciences
   253:383-&.
- Cameron, R. A. D. and L. M. Cook. 2012. Habitat and the shell polymorphism of
   *Cepaea nemoralis* (L.): interrogating the Evolution Megalab database. J
   Molluscan Stud 78:179-184.
- Cameron, R. A. D., L. M. Cook, and J. J. D. Greenwood. 2013. Change and stability
   in a steep morph-frequency cline in the snail *Cepaea nemoralis* (L.) over 43
   years. Biol J Linn Soc 108:473-483.
- Cassey, P., M. Honza, T. Grim, and M. E. Hauber. 2008. The modelling of avian
   visual perception predicts behavioural rejection responses to foreign egg
   colours. Biol Lett 4:515-517.
- Caves, E. M., P. A. Green, M. N. Zipple, S. Peters, S. Johnsen, and S. Nowicki.
   2018. Categorical perception of colour signals in a songbird. Nature.
- 515 Charlesworth, D. and B. Charlesworth. 1975a. Theoretical genetics of Batesian 516 mimicry I. Single-locus models. J Theor Biol 55:283-303.
- Charlesworth, D. and B. Charlesworth. 1975b. Theoretical Genetics of Batesian
   Mimicry II. Evolution of supergenes. J Theor Biol 55:305-324.
- 519 Cook, L. M. 2014. Morph frequency in British *Cepaea nemoralis*: what has changed 520 in half a century? J Molluscan Stud 80:43-46.
- Cook, L. M. 2017. Reflections on molluscan shell polymorphisms. Biol J Linn Soc
   121:717-730.
- 523 Cook, L. M., R. H. Cowie, and J. S. Jones. 1999. Change in morph frequency in the 524 snail *Cepaea nemoralis* on the Marlborough Downs. Heredity 82:336-342.
- Cook, L. M., B. S. Grant, I. J. Saccheri, and J. Mallet. 2012. Selective bird predation
   on the peppered moth: the last experiment of Michael Majerus. Biol Lett
   8:609-612.
- Cote, J., J. F. Le Galliard, J. M. Rossi, and P. S. Fitze. 2008. Environmentally
   induced changes in carotenoid-based coloration of female lizards: a comment
   on Vercken et al. J Evol Biol 21:1165-1172.
- Cuthill, I. C. 2006. Color perception. Pp. 3-40 *in* G. E. Hill, and K. J. McGraw, eds.
   Bird coloration. Cambridge (MA): Harvard University Press.
- Cuthill, I. C., W. L. Allen, K. Arbuckle, B. Caspers, G. Chaplin, M. E. Hauber, G. E.
  Hill, N. G. Jablonski, C. D. Jiggins, A. Kelber, J. Mappes, J. Marshall, R.
  Merrill, D. Osorio, R. Prum, N. W. Roberts, A. Roulin, H. M. Rowland, T. N.
  Sherratt, J. Skelhorn, M. P. Speed, M. Stevens, M. C. Stoddard, D. StuartFox, L. Talas, E. Tibbetts, and T. Caro. 2017. The biology of color. Science
  357

- 539 Dale, J., C. J. Dey, K. Delhey, B. Kempenaers, and M. Valcu. 2015. The effects of
  540 life history and sexual selection on male and female plumage colouration.
  541 Nature 527:367-370.
- 542 Davison, A. and B. Clarke. 2000. History or current selection? A molecular analysis
  543 of 'area effects' in the land snail *Cepaea nemoralis*. Proc R Soc Lond B Biol
  544 Sci 267:1399-1405.
- 545 Delhey, K., V. Delhey, B. Kempenaers, and A. Peters. 2015. A practical framework
   546 to analyze variation in animal colors using visual models. Behav Ecol 26:367 547 375.
- 548 Delhey, K., B. Szecsenyi, S. Nakagawa, and A. Peters. 2017. Conspicuous plumage 549 colours are highly variable. Proc R Soc Lond B Biol Sci 284.
- Endler, J. 1978. A predator's view of animal color patterns. Pp. 319-364 *in* M. K.
   Hecht, W. C. Steere, and B. Wallace, eds. Evolutionary biology. Plenum
   Press, New York.
- 553 Endler, J. A. 1990. On the measurement and classification of color in studies of 554 animal color patterns. Biol J Linn Soc 41:315-352.
- 555 Ford, E. B. 1964. Ecological Genetics. Methuen, London.
- 556 Franks, D. W. and G. S. Oxford. 2009. The evolution of exuberant visible 557 polymorphisms. Evolution 63:2697-2706.
- 558 Franks, D. W. and G. S. Oxford. 2011. The interrelationship between crypsis and 559 colour polymorphism. Ecol Lett 14:295-300.
- Franks, D. W. and G. S. Oxford. 2017. The co-evolution of anti-predator
   polymorphisms in sympatric populations. Biol J Linn Soc 122:729-737.
- Gautier, M., J. Yamaguchi, J. Foucaud, A. Loiseau, A. Ausset, B. Facon, B.
  Gschloessl, J. Lagnel, E. Loire, H. Parrinello, D. Severac, C. Lopez-Roques,
  C. Donnadieu, M. Manno, H. Berges, K. Gharbi, L. Lawson-Handley, L.-S.
  Zang, H. Vogel, A. Estoup, and B. Prud'homme. 2018. The genomic basis of
  colour pattern polymorphism in the harlequin ladybird. bioRxiv.
- Grindon, A. J. and A. Davison. 2013. Irish *Cepaea nemoralis* land snails have a
   cryptic Franco-Iberian origin that is most easily explained by the movements
   of Mesolithic humans. PLoS One 8:e65792.
- Hart, N. S. 2001. Variations in cone photoreceptor abundance and the visual ecology
   of birds. Journal of comparative physiology. A, Sensory, neural, and
   behavioral physiology 187:685-697.
- Hart, N. S., J. C. Partridge, I. C. Cuthill, and A. T. Bennett. 2000. Visual pigments, oil
  droplets, ocular media and cone photoreceptor distribution in two species of
  passerine bird: the blue tit (*Parus caeruleus* L.) and the blackbird (*Turdus merula* L.). J Comp Physiol A 186:375-387.

- Hoekstra, H. E. 2006. Genetics, development and evolution of adaptive pigmentation
   in vertebrates. Heredity 97:222-234.
- Holmes, I. A., M. R. Grundler, and A. R. D. Rabosky. 2017. Predator perspective
   drives geographic variation in frequency-dependent polymorphism. Am Nat
   190:E78-E93.
- Hugall, A. F. and D. Stuart-Fox. 2012. Accelerated speciation in colour-polymorphic
   birds. Nature 485:631-634.
- Jones, J. S., B. H. Leith, and P. Rawlings. 1977. Polymorphism in *Cepaea*: a problem with too many solutions? Annu Rev Ecol Syst 8:109-143.
- Joron, M., L. Frezal, R. T. Jones, N. L. Chamberlain, S. F. Lee, C. R. Haag, A.
  Whibley, M. Becuwe, S. W. Baxter, L. Ferguson, P. A. Wilkinson, C. Salazar,
  C. Davidson, R. Clark, M. A. Quail, H. Beasley, R. Glithero, C. Lloyd, S. Sims,
  M. C. Jones, J. Rogers, C. D. Jiggins, and R. H. ffrench-Constant. 2011.
  Chromosomal rearrangements maintain a polymorphic supergene controlling
  butterfly mimicry. Nature 477:203-206.
- Kang, C., M. Stevens, J. Y. Moon, S. I. Lee, and P. G. Jablonski. 2015. Camouflage
   through behavior in moths: the role of background matching and disruptive
   coloration. Behav Ecol 26:45-54.
- 595 Kerkvliet, J., T. de Boer, M. Schilthuizen, and K. Kraaijeveld. 2017. Candidate genes 596 for shell colour polymorphism in *Cepaea nemoralis*. Peerj 5:e3715.
- 597 Kopp, M. and J. Hermisson. 2006. The evolution of genetic architecture under 598 frequency-dependent disruptive selection. Evolution 60:1537-1550.
- Kunte, K., W. Zhang, A. Tenger-Trolander, D. H. Palmer, A. Martin, R. D. Reed, S.
  P. Mullen, and M. R. Kronforst. 2014. Doublesex is a mimicry supergene.
  Nature 507:229–232.
- Li, J. H., J. M. Cocker, J. Wright, M. A. Webster, M. McMullan, S. Dyer, D.
  Swarbreck, M. Caccamo, C. van Oosterhout, and P. M. Gilmartin. 2016.
  Genetic architecture and evolution of the S locus supergene in *Primula vulgaris*. Nat Plants 2.
- Llaurens, V., A. Whibley, and M. Joron. 2017. Genetic architecture and balancing
   selection: the life and death of differentiated variants. Mol Ecol 26:2430-2448.
- Maia, R., C. M. Eliason, P. P. Bitton, S. M. Doucet, and M. D. Shawkey. 2013. pavo:
   an R package for the analysis, visualization and organization of spectral data.
   Methods Ecol Evol 4:906-913.
- Mann, K. and D. J. Jackson. 2014. Characterization of the pigmented shell-forming
   proteome of the common grove snail *Cepaea nemoralis*. BMC Genomics 15.
- 613 McKinnon, J. S. and M. E. R. Pierotti. 2010. Colour polymorphism and correlated 614 characters: genetic mechanisms and evolution. Mol Ecol 19:5101-5125.

- McLean, C. A. and D. Stuart-Fox. 2014. Geographic variation in animal colour polymorphisms and its role in speciation. Biol Rev (Camb) 89:860-873.
- Montgomerie, R. 2006. Analysing Colors. Pp. 90-147 *in* G. E. Hill, and K. J. McGraw, eds. Bird Coloration. Cambridge (MA): Harvard University Press.
- Osorio, D., A. Miklosi, and Z. Gonda. 1999. Visual ecology and perception of coloration patterns by domestic chicks. Evol Ecol 13:673-689.
- Osorio, D. and M. Vorobyev. 2005. Photoreceptor spectral sensitivities in terrestrial
   animals: adaptations for luminance and colour vision. Proceedings of the
   Royal Society B-Biological Sciences 272:1745-1752.
- Ozgo, M., T. S. Liew, N. B. Webster, and M. Schilthuizen. 2017. Inferring
   microevolution from museum collections and resampling: lessons learned
   from *Cepaea*. Peerj 5.
- Ożgo, M. and M. Schilthuizen. 2012. Evolutionary change in *Cepaea nemoralis* shell
   colour over 43 years. Global Change Biology 18:74-81.

Paterson, J. E. and G. Blouin-Demers. 2017. Distinguishing discrete polymorphism
 from continuous variation in throat colour of tree lizards, *Urosaurus ornatus*.
 Biol J Linn Soc 121:72-81.

- Richards, P. M., M. M. Liu, N. Lowe, J. W. Davey, M. L. Blaxter, and A. Davison.
   2013. RAD-Seq derived markers flank the shell colour and banding loci of the
   *Cepaea nemoralis* supergene. Mol Ecol 22:3077-3089.
- San-Jose, L. M. and A. Roulin. 2017. Genomics of coloration in natural animal
   populations. Philos Trans R Soc Lond B Biol Sci 372.
- 637 Schilthuizen, M. 2013. Rapid, habitat-related evolution of land snail colour morphs on 638 reclaimed land. Heredity 110:247-252.
- Scrucca, L., M. Fop, T. B. Murphy, and A. E. Raftery. 2016. Mclust 5: clustering,
   classification and density estimation using Gaussian finite mixture models.
   The R Journal 8:205-233.
- Silvertown, J., L. Cook, R. Cameron, M. Dodd, K. McConway, J. Worthington, P.
  Skelton, C. Anton, O. Bossdorf, B. Baur, M. Schilthuizen, B. Fontaine, H.
  Sattmann, G. Bertorelle, M. Correia, C. Oliveira, B. Pokryszko, M. Ozgo, A.
  Stalazas, E. Gill, U. Rammul, P. Solymos, Z. Feher, and X. Juan. 2011.
  Citizen science reveals unexpected continental scale evolutionary change in a
  model organism. Plos One 6:e18927.
- 648 Sinervo, B. and C. M. Lively. 1996. The rock–paper–scissors game and the evolution 649 of alternative male strategies. Nature 380:240.
- Spottiswoode, C. N. and M. Stevens. 2010. Visual modeling shows that avian host
   parents use multiple visual cues in rejecting parasitic eggs. Proc Natl Acad
   Sci USA 107:8672-8676.

- Spottiswoode, C. N. and M. Stevens. 2011. How to evade a coevolving brood
   parasite: egg discrimination versus egg variability as host defences. Proc R
   Soc Biol Sci Ser B 278:3566-3573.
- 656 Staples-Browne, R. 1908. On the inheritance of colour in domestic pigeons, with 657 special reference to reversion. Proc Zool Soc Lond 78:67-104.
- Surmacki, A., A. Ozarowska-Nowicka, and Z. M. Rosin. 2013. Color polymorphism in
   a land snail *Cepaea nemoralis* (Pulmonata: Helicidae) as viewed by potential
   avian predators. Naturwissenschaften 100:533-540.
- Taylor, C. H., T. Reader, and F. Gilbert. 2016. Hoverflies are imperfect mimics of
   wasp colouration. Evol Ecol 30:567-581.
- Teasdale, L. C., M. Stevens, and D. Stuart-Fox. 2013. Discrete colour polymorphism
   in the tawny dragon lizard (*Ctenophorus decresii*) and differences in signal
   conspicuousness among morphs. J Evol Biol 26:1035-1046.
- Thompson, M. J. and C. D. Jiggins. 2014. Supergenes and their role in evolution.
   Heredity 113:1-8.
- Tripathi, N., M. Hoffmann, E. M. Willing, C. Lanz, D. Weigel, and C. Dreyer. 2009.
  Genetic linkage map of the guppy, *Poecilia reticulata*, and quantitative trait
  loci analysis of male size and colour variation. Proc R Soc Biol Sci Ser B
  276:2195-2208.
- van't Hof, A. E., P. Campagne, D. J. Rigden, C. J. Yung, J. Lingley, M. A. Quail, N.
  Hall, A. C. Darby, and I. J. Saccheri. 2016. The industrial melanism mutation
  in British peppered moths is a transposable element. Nature 534:102-105.
- Vercken, E., B. Sinervo, and J. Clobert. 2008. Colour variation in female common
  lizards: why we should speak of morphs, a reply to Cote et al. J Evol Biol
  21:1160-1164.
- Vorobyev, M. and D. Osorio. 1998. Receptor noise as a determinant of colour
   thresholds. Proc R Soc Lond B Biol Sci 265:351-358.
- Vorobyev, M., D. Osorio, A. T. D. Bennett, N. J. Marshall, and I. C. Cuthill. 1998.
   Tetrachromacy, oil droplets and bird plumage colours. J Comp Physiol A 183:621-633.
- 683 Wellenreuther, M., E. I. Svensson, and B. Hansson. 2014. Sexual selection and 684 genetic colour polymorphisms in animals. Mol Ecol 23:5398-5414.
- Wheldale, M. 1907. The inheritance of flower colour in *Antirrhinum majus*. Proc R
   Soc Biol Sci Ser B 79:288-305.
- 687 White, T. E. and D. J. Kemp. 2016. Color polymorphic lures target different visual 688 channels in prey. Evolution 70:1398-1408.
- 689 Williams, S. T. 2017. Molluscan shell colour. Biol Rev (Camb) 92:1039–1058.
- 690

**Table 1.** Comparison between human perceived colour categories and Mclust-defined groups. Shells that were scored the same are on the diagonal (in bold).Yellow and pink were most common, and so the absolute number of discordantscores was relatively low. Brown had by far the highest proportion of discordantscores.

		<b>Colour (human)</b> yellow pink brown total % total misclassifie						
Colour	yellow	495	44	0	539	8.2		
(Mclust)	pink brown	118	379	14	511	25.8		
		9	90	23	122	81.1		
	total	622	513	37				

**Table 2.** Results of likelihood ratio tests of the terms in binomial GLMMs of the effects of geographic variables and banding phenotype on the probability that a snail belonged to each of the three colour morphs. Significant p-values are in bold. The effects of modelling large-scale geographic variation in two ways are illustrated by the AIC values for the best model in which linear and quadratic effects of latitude and longitude were included (AICa), and the best model in which geographic region was included (AICb). All models include a random effect for site.

	Brov	vn		Pink			Yellow			
AlCa (df) AlCb (df)					865.2 (11) 875.4 (22)			850.2 (8) 872.09 (22)		
Term	df	χ2	р	df	χ2	р	df	χ2	р	
Latitude	1	1.747	0.186	1	0.218	0.641	1	2.62	0.106	
Longitude	1	1.572	0.21	1	1.382	0.24	1	0.001	0.986	
Latitude/2	1	2.019	0.155	1	1.832	< 0.001	1	4.732	0.03	
Longitude <sup>2</sup>	1	2.587	0.108	1	0.214	0.644	1	0.355	0.551	
Banding	2	10.751	0.005	2	0.286	0.867	2	3.056	0.217	
Latitude x longitude	1	0.239	0.625	1	0.0877	0.767	1	0.023	0.88	
Latitude x banding	2	1.411	0.494	2	6.345	0.042	2	4.972	0.083	
Longitude x banding	2	4.255	0.119	2	12.935	0.002	2	27.043	< 0.001	
Region	6	10.586	0.102	6	7.481	0.279	6	5.924	0.432	
Banding x region	12	12.824	0.382	12	34.564	0.001	12	31.448	0.002	

**Figure 1.** Sample sites across Europe, grouped by geographically contiguous regions. A (England, n=397), B (Ireland, n=144), C (North Spain and Pyrenees, n=112), D (North France, Belgium, Germany, n= 178), E (Scandinavia, n=77), F (Poland, n=126) and all others (n=138).



**Figure 2.** Axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space, shown from two different perspectives (see also Supplementary Movie 2). Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to Mclust classification of the shell, either yellow, pink or brown. Top: Variation along PC1 (87%) mainly represents differences in saturation between shells. PC2 (11%) shows relatively higher stimulation of L cones and lesser stimulation of M and S cones, and tends to separate brown from pink/yellow. Bottom: PC3 (2%) shows relatively high stimulation of the M cones compared to lesser stimulation of the S and L cones, and tends to separate yellow from pink and pink from brown.



**Figure 3.** Interquartile ranges of the average normalised reflectance spectra for the principal component axes shown in Figure 2. These plots confirm that variation on PC1 mainly represents differences in saturation between shells; PC2 represents relatively higher stimulation of L cones and lesser stimulation of M and S cones; PC3 represents relatively high stimulation of the M cones compared to lesser stimulation of the S and L cones.



**Figure 4.** Scatterplot and associated density plot, showing variation along three principal component axes. Units are in JNDs. Points are coloured according to Mclust classification of the shell, either yellow, pink or brown. Top: PC1 versus PC2. Bottom: PC2 versus PC3.



Figure 5. Scaled effects of latitude on the proportion of pink shells (top), and

longitude on banding and proportion of pink shells.



**Figure 6.** Scaled effects of latitude on the proportion of yellow shells (top), and longitude on banding and proportion of yellow shells.



Supplementary Figure 1. Boxplot showing extent of achromatic variation in Mclust-

defined colour morphs of Cepaea nemoralis.



**Supplementary Movie 1.** Animation showing axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space. Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to human-scoring of the shell, either yellow, pink or brown.

**Supplementary Movie 2.** Animation showing axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space. Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to Mclust classification of the shell, either yellow, pink or brown.